federal register

WEDNESDAY, MAY 19, 1971
WASHINGTON, D.C.
Volume 36 ■ Number 97

PART II

DEPARTMENT OF AGRICULTURE

Agricultural Research Service

■

National Poultry and Turkey Improvement Plans and Auxiliary Provisions

■

Notice of Proposed Rule Making

DEPARTMENT OF AGRICULTURE

Agricultural Research Service

[9 CFR Parts 145, 147]

NATIONAL POULTRY AND TURKEY IMPROVEMENT PLANS AND AUXILIARY PROVISIONS

Notice of Proposed Rule Making

Notice is hereby given, under the administrative procedure provisions of 5 U.S.C. 553, that the Department of Agriculture has under consideration proposed amendments of the National Poultry and Turkey Improvement Plans and Auxiliary Provisions recommended by the 1970 Conference of representatives of the State agencies cooperating in the administration of the Plans, and by the General Conference Committee, and that, pursuant to section 101(b) of the Department of Agriculture Organic Act of 1944, as amended (7 U.S.C. 429), it is proposed to revise Parts 145, 146, and 147 of Title 9, Chapter I, Subchapter F, Code of Federal Regulations, to incorporate such recommended amendments and to make incidental changes for clarity and consistency.

The recommendations included the general revision of Parts 145, 146, and 147 into one Plan, with the consolidation of the general provisions applicable to all classes of fowl and providing for special provisions applicable to problems and conditions peculiar to particular classes of fowl. To comply with these recommendations, it is proposed to consolidate the general requirements for participation and administration of the National Poultry Improvement Plan (Part 145) and the National Turkey Improvement Plan (Part 146) in Subpart A, and to transfer the special provisions for four classes of fowl: (1) Egg type chickens, (2) meat type chickens, (3) turkeys, and (4) waterfowl, exhibition poultry, and game birds to Subparts B, C, D, and E, respectively, of Part 145, which would be designated as National Poultry Improvement Plan. The procedures for conducting random sample performance tests would be transferred to Part 147, which would be designated as Auxiliary Provisions of the National Poultry Improvement Plan. Said Subchapter F, Poultry Improvement, would be revised to read as follows:

PART 145—NATIONAL POULTRY IMPROVEMENT PLAN

Subpart A—General Provisions

Sec.
145.1 Definitions.
145.2 Administration.
145.3 Participation.
145.4 General provisions for all participants.
145.5 Specific provisions for participating flocks.
145.6 Specific provisions for participating hatcheries.
145.7 Specific provisions for participating dealers.
145.8 Terminology and classification; general.
145.9 Terminology and classification; hatcheries and dealers.
145.10 Terminology and classification; flocks and products.
145.11 Supervision.
145.12 Inspections.
145.13 Debarment from participation.
145.14 Blood testing.

Subpart B—Special Provisions for Egg Type Chicken Breeding Flocks and Products

145.21 Definitions.
145.22 Participation.
145.23 Terminology and classification.

Subpart C—Special Provisions for Meat Type Chicken Breeding Flocks and Products

145.31 Definitions.
145.32 Participation.
145.33 Terminology and classification.

Subpart D—Special Provisions for Turkey Breeding Flocks and Products

145.41 Definitions.
145.42 Participation.
145.43 Terminology and classification.

Subpart E—Special Provisions for Waterfowl, Exhibition Poultry, and Game Bird Breeding Flocks and Products

145.51 Definitions.
145.52 Participation.
145.53 Terminology and classification.

Subpart A—General Provisions

§ 145.1 Definitions.

Words used in this part in the singular form shall be deemed to import the plural, and vice versa, as the case may demand. Except where the context otherwise requires, for the purposes of this part the following terms shall be construed, respectively, to mean:

(a) *Plan.* The provisions of the National Poultry Improvement Plan contained in this part.

(b) *Person.* A natural person, firm, or corporation.

(c) *Department.* The U.S. Department of Agriculture.

(d) *ASR Division.* The Animal Science Research Division of the Agricultural Research Service of the Department.

(e) *State.* Any State, the District of Columbia, or Puert Rico.

(f) *Official State Agency.* The State authority recognized by the Department to cooperate in the administration of the Plan.

(g) *State Inspector.* Any person employed or authorized under § 145.11(b) to perform functions under this part.

(h) *Authorized Agent.* Any person designated under § 145.11(a) to perform functions under this part .

(i) *Affiliated flockowner.* A flockowner who is participating in the Plan through an agreement with a participating hatchery.

(j) *Flock*—(1) *As applied to breeding.* All poultry of one kind of mating (breed and variety or combination of stocks) and of one classification on one farm;

(2) *As applied to disease control.* All of the poultry on one farm except that, at the discretion of the Official State Agency, any group of poultry which is segregated from another group and has been so segregated for a period of at least 21 days may be considered as a separate flock.

(k) *Hatchery.* Hatchery equipment on one premises operated or controlled by any person for the production of baby poultry.

(l) *Poultry.* Domesticated fowl, including chickens, turkeys, waterfowl, and game birds, except doves and pigeons, which are bred for the primary purpose of producing eggs or meat.

(m) *Domesticated.* Propagated and maintained under the control of a person.

(n) *Products.* Poultry breeding stock and hatching eggs, baby poultry, and started poultry.

(o) *Baby poultry.* Newly hatched poultry (chicks, poults, ducklings, goslings, keets, etc.) that have not been fed or watered.

(p) *Started poultry.* Young poultry (chicks, pullets, cockrels, capons, poults, ducklings, goslings, keets, etc.) that have been fed and watered and are less than 6 months of age.

(q) *Strain.* Poultry breeding stock bearing a given name produced by a breeder through at least five generations of closed flock breeding.

(r) *Stock.* A term used to identify the progeny of a specific breeding combination within a species of poultry. These breeding combinations may include pure strains, strain crosses, breed crosses, or combinations thereof.

(s) *Primary breeding flock.* A flock composed of one or more generations that is maintained for the purpose of establishing, continuing, or improving parent lines.

(t) *Multiplier breeding flock.* A flock that originated from a primary breeding flock and is intended for the production of hatching eggs used for the purpose of producing progeny for commercial egg or meat production or for other nonbreeding purposes.

(u) *Trade name or number.* A name or number compatible with State or Federal laws and regulations applied to a specific stock or product thereof.

(v) *Franchise breeder.* A breeder who normally sells products under a specific strain or trade name and who authorizes other hatcheries to produce and sell products under this same strain or trade name.

(w) *Franchise hatchery.* A hatchery which has been authorized by a franchise breeder to produce and sell products under the breeder's strain or trade name.

(x) *Pullorum Disease or Pullorum.* A disease of poultry caused by Salmonella pullorum.

(y) *Fowl Typhoid or Typhoid.* A disease of poultry caused by Salmonella gallinarum.

(z) *S. Typhimurium Infection or Typhimurium.* A disease of poultry caused by Salmonella typhimurium or S. typhimurium var. copenhagen.

§ 145.2 Administration.

(a) The Department cooperates through a Memorandum of Understanding with Official State Agencies in the administration of the Plan.

(b) The Official State Agency shall carry out the administration of the Plan within the State according to the applicable provisions of the Plan and the Memorandum of Understanding. An Official State Agency may accept for participation an affiliated flock located in another State under a mutual understanding and agreement, in writing, between the two Official State Agencies regarding conditions of participation and supervision.

(c) The Official State Agency of any State may, except as limited by § 145.3(d), adopt regulations applicable to the administration of the Plan in such State further defining the provisions of the Plan or establishing higher standards compatible with the Plan.

§ 145.3 Participation.

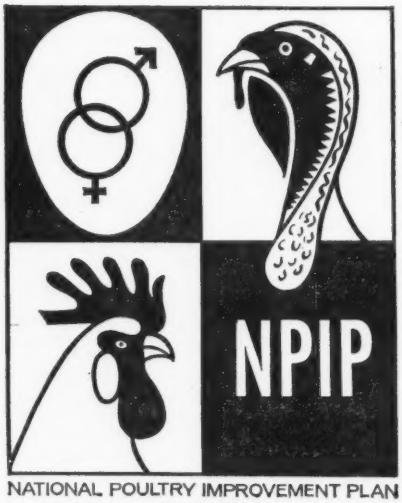
(a) Any person producing or dealing in poultry products may participate in the Plan when he has demonstrated, to the satisfaction of the Official State Agency, that his facilities, personnel, and practices are adequate for carrying out the applicable provisions of the Plan, and has signed an agreement with the Official State Agency to comply with the general and the applicable specific provisions of the Plan and any regulations of the Official State Agency under § 145.2. Affiliated flockowners may participate without signing an agreement with the Official State Agency.

(b) Each participant shall comply with the Plan throughout the operating year of the Official State Agency, or until released by such Agency.

(c) A participant in any State shall participate with all of his poultry hatching egg supply flocks and hatchery operations within such State.

(d) No person shall be compelled by the Official State Agency to qualify products for any of the other classifications described in § 145.10 as a condition of qualification for the U.S. Pullorum-Typhoid Clean classification.

(e) Participation in the Plan shall entitle the participant to use the Plan emblem reproduced below:



NATIONAL POULTRY IMPROVEMENT PLAN

FIGURE 1.

§ 145.4 General provisions for all participants.

(a) Records of purchases and sales and the identity of products handled shall be maintained in a manner satisfactory to the Official State Agency.

(b) Products, records of sales, and purchase of products, and material used to advertise products shall be subject to inspection by the Official State Agency at any time.

(c) Advertising must be in accordance with the Plan, and applicable rules and regulations of the Official State Agency and the Federal Trade Commission. A participant advertising products as being of any official classification may include in his advertising reference to associated or franchised hatcheries only when such hatcheries produce the same kind of products of the same classification.

(d) Participants may not buy or receive for any purpose products from nonparticipants, or sell products of nonparticipants, except with the permission of the Official State Agency for use in breeding flocks or for experimental purposes.

(e) Each shipment of products to points outside the United States and its territories and possessions shall be accompanied by a properly executed NPIP Form 15F, Report of Sales of Hatching Eggs, Chicks, and Poults (For Shipment Outside the United States).

§ 145.5 Specific provisions for participating flocks.

(a) Poultry equipment, and poultry houses, and the land in the immediate vicinity thereof, shall be kept in sanitary condition, and the participating flock, its eggs, and all equipment used in connection with the flock shall be separated from nonparticipating flocks, in a manner acceptable to the Official State Agency. The procedures outlined in §§ 147.21 and 147.22 (a) and (e) of this chapter shall be considered as a guide in determining compliance with this provision.

(b) All flocks shall consist of healthy, normal individuals characteristic of the breed, variety, cross, or other combination which they are stated to represent.

(c) A flock shall be deemed to be a participating flock at any time only if, within the past 12 months, it has qualified for the U.S. Pullorum-Typhoid Clean classification, as prescribed in Subpart B, C, D, or E of this part.

(d) Each bird shall be identified with a sealed and numbered band obtained through or approved by the Official State Agency: *Provided,* That exception may be made at the discretion of the Official State Agency.

§ 145.6 Specific provisions for participating hatcheries.

(a) Hatcheries, including brooder rooms, shall be kept in sanitary condition, acceptable to the Official State Agency. The procedures outlined in §§ 147.22 through 147.25 of this chapter shall be considered as a guide in determining compliance with this provision. The minimum requirements with respect to sanitation shall include the following:

(1) Incubator walls, floors, and trays shall be kept free from broken eggs and eggshells.

(2) Tops of incubators and hatchers shall be kept clean (not used for storage).

(3) Entire hatchery, including sales room, shall be kept in a neat, orderly condition, and free from accumulated dust.

(4) Hatchery residue, such as eggshells, infertile eggs, and dead germs, shall be disposed of promptly and in a manner satisfactory to the Official State Agency.

(5) Hatchers and hatching trays shall be cleaned and fumigated or disinfected after each hatch, preferably using the procedures outlined in §§ 147.24(b) and 147.25(e) of this chapter.

(b) A hatchery which keeps started poultry must keep such poultry separated from the incubator room in a manner satisfactory to the Official State Agency.

(c) All baby and started poultry offered for sale under Plan terminology shall be normal and typical of the breed, variety, cross, or other combination represented.

(d) Eggs incubated shall be sound in shell, typical for the breed, variety, strain, or cross thereof, and reasonably uniform in shape. Hatching eggs shall be trayed and the baby poultry boxed with a view to uniformity of size.

(e) All hatcheries within a State which are operated under the ownership or management of the same person or persons or related corporations, or in which the same person or persons have a substantial financial interest as partners or otherwise, shall participate in the Plan if any of them are to participate.

§ 145.7 Specific provisions for participating dealers.

Dealers in poultry breeding stock, hatching eggs, or baby or started poultry shall comply with all provisions in this part which apply to their operations.

§ 145.8 Terminology and classification; general.

(a) The official classification terms defined in §§ 145.9 and 145.10 and the various designs illustrative of the official classifications reproduced in § 145.10 may be used only by participants and to describe products that have met all the specific requirements of such classifications.

(b) Products produced under the Plan shall lose their identity under Plan terminology when they are purchased for resale by or consigned to nonparticipants.

(c) Participating flocks, their eggs, and the baby and started poultry produced from them may be designated by their strain or trade name. When a

breeder's trade name or strain designation is used, the participant shall be able by records to substantiate that the products so designated are from flocks that are composed of either birds hatched from eggs produced under the direct supervision of the breeder of such strain, or stock multiplied by persons designated and so reported by the breeder to each Official State Agency concerned.

§ 145.9 Terminology and classification; hatcheries and dealers.

Participating hatcheries and dealers shall be designated as "National Plan Hatchery" and "National Plan Dealer", respectively. Each participating hatchery or dealer may be assigned a permanent approval number by the ASR Division. This number may appear on each invoice and shipping label for each separate sale of products. The approval number shall be withdrawn when the hatchery or dealer no longer qualifies for participation in the Plan. All Official State Agencies shall be notified by the ASR Division of additions, withdrawals, and changes in classification.

§ 145.10 Terminology and classification; flocks and products.

Participating flocks, and the products produced from them, which have met the respective requirements specified in Subpart B, C, D, or E of this part may be designated by the following terms or illustrative designs:

(a) *U.S. Record of Performance.* (See § 145.23(a).)

FIGURE 2.

(b) *U.S. Performance Tested Parent Stock.* (See §§ 145.23(b) and 145.33(a).)

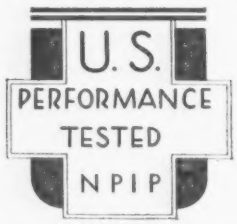


FIGURE 3.

(c) *U.S. Certified for Eggs.* (See § 145.23(c).)

FIGURE 4.

(d) *U.S. Certified for Meat.* (See § 145.33(b).)

FIGURE 5.

(e) *U.S. Approved.* (See §§ 145.23(d), 145.33(c), 145.43(a), and 145.53(a).)

FIGURE 6.

(f) *U.S. Pullorum-Typhoid Clean.* (See §§ 145.23(e), 145.33(d), 145.43(b), and 145.53(b).)

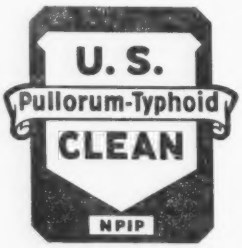


FIGURE 7.

(g) *U.S. M. Gallisepticum Clean.* (See §§ 145.23(f), 145.33(e), 145.43(c), and 145.53(c).)

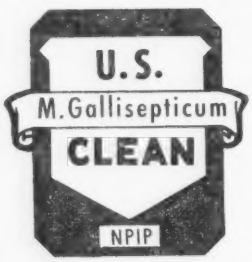


FIGURE 8.

(h) *U.S. Typhimurium Controlled.* (See § 145.43(d).)

FIGURE 9.

§ 145.11 Supervision.

(a) The Official State Agency may designate qualified persons as Authorized Agents to do the blood collecting and blood testing provided for in §§ 145.5 and 145.14, and the selecting required for the U.S. approved classification provided for in § 145.10(e).

(b) The Official State Agency shall employ or authorize qualified persons as State Inspectors to perform or supervise the performance of the selecting and testing of participating flocks, and to perform the official inspections necessary to verify compliance with the requirements of the Plan.

§ 145.12 Inspections.

(a) Each participating hatchery shall be inspected a sufficient number of times each year to satisfy the Official State Agency that the operations of the hatchery are in compliance with the provisions of the Plan.

(b) Each year at least 15 percent of the flocks selected or tested by each Authorized Agent shall be inspected by a State Inspector. This must include the inspection of some flocks of each hatchery. Each flock inspection shall include the examination of a sufficient number of males and females and, in flocks qualified for participation by the whole-blood test, the blood testing of a sufficient number of birds to determine whether the work of the Authorized Agent was satisfactory and that the flock is qualified for participation.

§ 145.13 Debarment from participation.

Noncompliance with the provisions of the Plan, or regulations of the Official State Agency under § 145.2, not corrected within the time specified by the Official State Agency, shall be grounds for the Official State Agency to bar a participant from further participation for a period to be determined in each case by the Official State Agency. Such action shall not be taken until a thorough investigation has been made by the Official State Agency and the participant has been given an opportunity for a hearing.

§ 145.14 Blood Testing.

Blood samples for official tests shall be drawn by an Authorized Agent or State Inspector and tested by an authorized laboratory, except that the stained-antigen, rapid whole-blood test for pullorum-typhoid may be conducted by an Authorized Agent or State Inspector.

(a) *For Salmonella.* (1) The Official blood tests for pullorum-typhoid shall be the standard tube agglutination test or the rapid serum test for all classes of poultry, or the stained-antigen, rapid whole-blood test for all classes of poultry except turkeys. The recommended procedures for conducting such tests are described in §§ 147.1, 147.2, and 147.3 of this chapter. Each lot of antigen used for the whole-blood test shall be approved by the Department and shall be of the polyvalent type.

(2) The official blood test for typhimurium shall be the standard tube agglutination test as described in § 147.4 of this chapter: *Provided,* That, if the following conditions are fulfilled, the tests for pullorum-typhoid and typhimurium may be combined:

(i) The flock is located in a State where an adequate surveillance program for pullorum-typhoid and typhimurium exists;

(ii) A single combination antigen composed of equal quantities of pullorum antigen and typhimurium antigen is used in a screening test in accordance with the procedures described in § 147.1 of this chapter;

(iii) All serum showing suspicious and positive reactions to the combination antigen are reset with individual antigens. Final determination of the status of each flock is determined by bacteriological examination of representative birds showing suspicious or positive reactions;

(iv) If the flock is found to be infected with S. pullorum, S. gallinarum, or S. typhimurium on the basis of bacteriological examinations, retests of the flock are made with separate antigens (pullorum and typhimurium antigens) until the flock is qualified or eliminated.

(3) There shall be an interval of at least 21 days between any official blood test and any previous test with Salmonella antigen.

(4) Turkeys must be more than 4 months of age and chickens and other poultry more than 5 months of age when tested.

(5) The official blood test shall include the testing of a sample of blood from each bird in the flock: *Provided,* That under specified conditions (see applicable provisions of §§ 145.23, 145.33, 145.43, and 145.53) the testing of a portion or sample of the birds may be used in lieu of testing each bird. When partial or sample testing is specified, the birds tested shall be a random or representative sample drawn on a pro rata basis from all pens or units of the flock. When reactors are found in any flock, or S. pullorum or S. gallinarum isolations are made from baby poultry or fluff samples, the flock may qualify for participation with two consecutive official negative tests. Qualification of this flock, or any other flock on the same premises during the next 12 months, shall be based on the testing of all birds, except that when the flock involved is turkeys, the period during which all birds must be tested shall be 2 years. Such testing shall be conducted by or directly supervised by a State Inspector.

(6) All domesticated fowl, except waterfowl, on the farm of the participant shall either be properly tested to meet the same standards as the participating flock or these birds and their eggs shall be separated from the participating flock and its eggs.

(7) All tests with Salmonella antigens of flocks participating in or candidates for participation in the Plan shall be reported to the Official State Agency within 10 days following the completion of such tests. All reactors shall be considered in determining the classification of the flock.

(8) (i) Reactors shall be submitted to a laboratory for autopsy and bacteriological examination. The laboratory and the number of reactors to be submitted shall be designated by the Official State Agency: *Provided,* That for turkey flocks, all reactors, if four or less, and a minimum of four, if there are more than four, shall be submitted. The recommended minimum procedure for bacteriological examination is described in § 147.11 of this chapter.

(ii) When reactors are submitted within 10 days from date of reading the test and the bacteriological examination fails to demonstrate infection of the serotype for which the test was conducted, the flock shall be deemed to have had no reactors to the specified test. If other members of the Salmonella group or paracolons are isolated, the Official State Agency may disqualify the flock for participation, or require such other action as is deemed necessary with respect to the infection.

(9) Any drug, for which there is scientific evidence of masking the test reaction or hindering the bacteriological recovery of Salmonella organisms, shall not be fed or administered to poultry within 3 weeks prior to a test or bacteriological examination upon which a Salmonella classification is based.

(10) When suitable evidence, as determined by the Official State Agency or the State Animal Disease Control Official, indicates that baby or started poultry produced by participating hatcheries are infected with organisms for which the parent flock received an official control classification and this evidence indicates egg transmission, the Official State Agency may, at its discretion, require additional testing of the flock involved. If infection is found in the parent flock, its classification shall be suspended until the flock is requalified under the requirements for the classification. Furthermore, the Official State Agency may require that the hatching eggs from such flocks be removed from the incubator and destroyed prior to hatching. When Salmonella or Arizona organisms are isolated from a specimen which originated in a participating hatchery, the Official State Agency shall attempt to locate and eliminate the source of the infection. The results of the investigation shall be reported by the Official State Agency to the ASR Division.

(b) *For M. gallisepticum.* (1) The official blood test for M. gallisepticum shall be either the serum plate agglutination test, the tube agglutination test, the hemagglutination inhibition (HI) test, or a combination of two or more of these tests. The HI test shall be used to confirm the results of other serological tests.

(2) The tests shall be conducted using M. gallisepticum antigen approved by the Department or the Official State Agency and shall be performed in accordance with the recommendations of the producer of the antigen.

(3) When reactors are submitted to a laboratory as prescribed by the Official State Agency, the following criteria shall be used to determine if the flock is negative for M. gallisepticum: (i) Active air sac lesions; (ii) recovery of M. gallisepticum; and (iii) supplemental serological tests. If all of these tests are negative, the flock shall be deemed to have had no M. gallisipticum reactors. If M. gallisepticum is recovered, the the flock shall be considered infected. If any of the other tests (subdivision (i) or (iii) of this subparagraph) is positive, the flock shall be considered suspicious, and additional laboratory tests shall be conducted before the final disposition of the flock is determined.

Subpart B—Special Provisions for Egg Type Chicken Breeding Flocks and Products

§ 145.21 Definitions.

Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:

(a) *Egg type chicken breeding flocks.* Flocks that are composed of stock that has been developed for egg production and are maintained for the principal purpose of producing chicks for the ultimate production of eggs for human consumption.

(b) *Baby chicks.* Chicks that have not been fed or watered.

(c) *Started chickens.* Young chickens (chicks, pullets, cockerels, capons) which have been fed and watered and are less than 6 months of age.

(d) *USROP or ROP.* U.S. Record of Performance.

(e) *ROP Supervisor.* The person employed or authorized to perform functions under § 145.23(a).

§ 145.22 Participation.

Participating flocks of egg type chickens, and the eggs and chicks produced from them, shall comply with the applicable general provisions of Subpart A of this part and the special provisions of this Subpart B.

(a) The minimum weight of hatching eggs sold shall be $1^{11}/_{12}$ ounces each, except as otherwise specified by the purchaser of the eggs.

(b) Mediterranean breed eggs shall be reasonably free from tints.

(c) All eggs should be fumigated at the hatchery as described in § 147.25 (a) or (b) or (c) of this chapter. All eggs should be refumigated after transfer to the hatcher as described in § 147.25(d) of this chapter.

(d) Started chickens shall lose their identity under Plan terminology when not maintained by Plan participants under the conditions prescribed in § 145.5(a).

§ 145.23 Terminology and classification.

Participating flocks, and the eggs and chicks produced from them, which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in § 145.10:

(a) *U.S. Record of Performance.* The ROP classification may be attained

through trapnesting and pedigree breeding under the supervision of an Official State Agency.

(1) Females may qualify as ROP females when they have been trapnested for a period of at least 6 months, and records of egg production and egg weight are maintained by the breeder.

(2) A male may qualify as an ROP male when his pedigree record, maintained by the breeder, shows he was produced from a single-male mating of an ROP female and the son of an ROP female.

(3) When products are sold or offered for sale under the ROP classification, the breeder shall have on file evidence that such products are from single-male matings of ROP males and ROP females.

(4) The ROP Supervisor shall represent the Official State Agency in its supervision of ROP participation. He shall visit and inspect the work of each breeder periodically.

(b) *U.S. Performance Tested Parent Stock.* The Performance Tested Parent Stock classification may be attained by stock represented by an entry in a random sample egg production test for which the records have been included in a combined summary published by the ASR Division. (See §§ 147.31, 147.32, and 147.34 of this chapter.) Application for the classification shall be made to the Official State Agency by the breeder of the parent stock. Such application, if acceptable to the Official State Agency, shall be submitted to the ASR Division.

(1) A stock may qualify as Performance Tested Parent Stock for egg production when the regressed mean of the stock for income above feed and chick cost per pullet housed, as published in the combined summary, exceeds the overall mean for all entries in all tests or is not significantly different, at the 5-percent level of probability, from the stock with the highest regressed mean.

(2) Qualification for the U.S. Performance Tested Parent Stock classification shall be determined by the ASR Division from records published in the combined summary, and that Division shall notify each applicant and his Official State Agency of his qualification or failure to qualify.

(3) Stock classified as Performance Tested Parent Stock may retain that classification for 1 year after qualification, provided the stock is maintained under the supervision of the qualifying breeder and is mated in the same combination, and for 1 more year when, in addition, the stock has been continuously represented by an entry for which the test results have been included in the combined summary. When the entry on which qualification is based is the progeny of a combination of two stocks which are distributed commercially under different strain or trade names, each stock may be designated as Performance Test Parent Stock, but this interrelationship shall be specified when the classification of either stock is referred to in advertising or certification.

(4) When products are sold or offered for sale as Performance Tested Parent Stock, the breeder shall be able to substantiate by records filed with the Official State Agency that such products are qualified for this classification.

(c) *U.S. Certified for Eggs.* All males ROP or all males and females from Performance Tested Parent Stock for egg production mated in the same combination as used in the qualifying parent flock.

(d) *U.S. Approved.* All males and females selected by Authorized Agents or State Inspectors according to standards prescribed by the Official State Agency or the State College of Agriculture.

(e) *U.S. Pullorum-Typhoid Clean.* A flock in which freedom from pullorum and typhoid has been demonstrated by one of the following criteria (See § 145.14 relating to the official blood test.):

(1) It has been officially blood tested within the past 12 months with no reactors.

(2) It is a multiplier breeding flock meeting the following specifications:

(i) The flock is located in a State where all persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is isolated.

(ii) The flock is composed entirely of birds that originated from U.S. Pullorum-Typhoid Clean primary breeding flocks or from flocks that met equivalent requirements under official supervision; and

(iii) A sample comprised of at least 25 percent of the birds in the flock has been officially blood tested within the past 12 months with no reactors, or its progeny has been subjected to an approved 10-day chick mortality bacteriological examination monitoring program and bacteriological examination of a fluff sample from selected hatches as prescribed by the Official State Agency: *Provided,* That when the blood testing procedure is used, the percentage of the flock included in the sample may be reduced by 5 percentage points following each year in which there is no evidence of infection on the premises: *And provided further,* That the sample tested for the qualification of a flock under this subparagraph shall include at least 500 birds the first year, 400 the second year, 300 the third year, 200 the fourth year, and 100 the fifth year.

(3) It is a multiplier breeding flock composed entirely of birds that originated from U.S. Pullorum-Typhoid Clean primary breeding flocks or from flocks that met equivalent requirements under official supervision, and is located in a State in which it has been determined by the ASR Division that:

(i) All hatcheries, except turkey hatcheries, within the State are qualified as "National Plan Hatcheries" or have met equivalent requirements for pullorum-typhoid control under official supervision.

(ii) All hatchey supply flocks, except turkey flocks, within the State, are qualified as U.S. Pullorum-Typhoid Clean or have met equivalent requirements for pullorum-typhoid control under official supervision: *Provided,* That if other domesticated fowl are maintained on the same premises as the participating flock, freedom from pullorum-typhoid infection shall be demonstrated by an official blood test of each of these fowl;

(iii) All shipments of product other than U.S. Pullorum-Typhoid Clean, or equivalent, into the State are prohibited;

(iv) All persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is isolated;

(v) All reports of S. pullorum or S. gallinarum isolations from poultry are promptly followed by an investigation by the Official State Agency to determine the origin of the infection;

(vi) All flocks found to be infected with pullorum or typhoid are quarantined until marketed or destroyed under the supervision of the Official State Agency, or until subsequently blood tested, following the procedure for reacting flocks as contained in § 145.14(a)(5), and all birds fail to demonstrate pullorum or typhoid infection;

(vii) All poultry, except turkeys, going to public exhibition come from U.S. Pullorum-Typhoid Clean or equivalent flocks, or have had a negative pullorum-typhoid test within 90 days of going to public exhibition;

(viii) Discontinuation of any of the conditions or procedures described in subdivisions (i), (ii), (iii), (iv), (v), (vi), and (vii) of this subparagraph, or the occurrence of repeated outbreaks of pullorum or typhoid in poultry breeding flocks, other than turkey flocks, within or originating within the State shall be grounds for revocation of the determination. Such action shall not be taken until a thorough investigation has been made by the ASR Division and the Official State Agency has been given an opportunity for a hearing.

(4) It is a multiplier breeding flock located in a State which has been determined to be in compliance with the provisions of § 145.23(e)(3), and in which pullorum disease or fowl typhoid is not known to exist nor to have existed in hatchery supply flocks, other than turkey flocks, within the State during the preceding 24 months.

(5) It is a primary breeding flock located in a State determined to be in compliance with the provision of § 145.23(e)(4), and in which a sample of 300 birds from flocks of more than 300, and each bird in flocks of 300 or less, has been officially tested for pullorum-typhoid within the past 12 months with no reactors: *Provided,* That a bacteriological examination monitoring program acceptable to the Official State Agency and approved by the ASR Division may be used in lieu of blood testing.

(f) *U.S. M. Gallisepticum Clean.* (1) A flock maintained in compliance with

the provisions of § 147.26 of this chapter and in which freedom from M. gallisepticum has been demonstrated by one of the following procedures:

(i) All birds have been tested as provided in § 145.14(b) when more than 5 months of age: *Provided,* That to retain this classification a random sample of at least 10 percent of the flock shall be tested at intervals of not more than 60 days; or

(ii) It is a multiplier breeding flock which originated as U.S. M. Gallisepticum Clean chicks from primary breeding flocks, and samples comprising at least 10 percent of the birds in the flock have been tested, as provided in § 145.14(b), twice between the ages of 8 weeks and 22 weeks, with an interval of not less than 60 days between the two tests: *Provided,* That to retain this classification, the flock shall be subjected to one of the following procedures:

(*a*) At intervals of not more than 90 days, a random sample of at least 5 percent of the flock shall be tested; or

(*b*) At intervals of not more than 30 days, a sample of 25 cull chicks produced from the flock shall be subjected to approved laboratory procedures for the detection and recovery of M. gallisepticum; or

(*c*) At intervals of not more than 60 days, serum samples obtained from at least 100-day-old chicks produced from the flock shall be examined for M. gallisepticum antibodies.

(2) A participant handling U.S. M. Gallisepticum Clean products shall keep these products separate from other products in a manner satisfactory to the Official State Agency: *Provided,* That U.S. M. Gallisepticum Clean chicks from primary breeding flocks shall be produced in incubators and hatchers in which only eggs from flocks qualified under subdivision (i) of subparagraph (1) of this paragraph are set.

(3) U.S. M. Gallisepticum Clean chicks shall be boxed in clean boxes and delivered in clean, disinfected trucks.

Subpart C—Special Provisions for Meat Type Chicken Breeding Flocks and Products

§ 145.31 Definitions.

Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:

(a) *Meat type chicken breeding flocks.* Flocks that are composed of stock that has been developed for meat production and are maintained for the principal purpose of producing chicks for the ultimate production of meat.

(b) *Baby chicks.* Chicks that have not been fed or watered.

(c) *Started chickens.* Young chickens (chicks, pullets, cockerels, capons) which have been fed and watered and are less than 6 months of age.

§ 145.32 Participation.

Participating flocks of meat-type chickens, and the eggs, and chicks produced from them, shall comply with the applicable general provisions of Subpart A of this part and the special provisions of this Subpart C.

(a) The minimum weight of hatching eggs sold shall be $1^{10}/_{12}$ ounce each, except as otherwise specified by the purchaser of the eggs.

(b) All eggs should be fumigated at the hatchery as described in § 147.25 (a) or (b) or (c) of this chapter. All eggs should be refumigated after transfer to the hatcher as described in § 147.25(d) of this chapter.

(c) Started chickens shall lose their identity under Plan terminology when not maintained by Plan participants under the conditions prescribed in § 145.5(a).

§ 145.33 Terminology and classification.

Participating flocks, and the eggs and chicks produced from them, which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in § 145.10:

(a) *U.S. Performance Tested Parent Stock.* The Performance Tested Parent Stock classification may be attained by a stock represented by an entry in a random sample meat production test for which the records have been included in a combined summary published by the ASR Division. (See §§ 147.31, 147.33, and 147.34 of this chapter.) Application for the classification shall be made to the Official State Agency by the breeder of the parent stock. Such application, if acceptable to the Official State Agency, shall be submitted to the ASR Division.

(1) A stock may qualify as Performance Tested Parent Stock for meat production when the regressed mean of the stock for rate of growth (average live weight at completion of test) and for rate of egg production on a hen-housed basis, as published in the combined summary, exceeds the overall mean for all entries in all tests or is not significantly different, at the 5-percent level of probability, from the stock with the highest regressed mean.

(2) Qualification for the U.S. Performance Tested Parent Stock classification shall be determined by the ASR Division from records published in the combined summary, and that Division shall notify each applicant and his Official State Agency of his qualification or failure to qualify.

(3) Stock classified as Performance Tested Parent Stock may retain that classification for 1 year after qualification, provided the stock is maintained under the supervision of the qualifying breeder and is mated in the same combination, and for 1 more year when, in addition, the stock has been continuously represented by an entry for which the test results have been included in the combined summary. When the entry on which qualification is based is the progeny of a combination of two stocks which are distributed commercially under different strain or trade names, each stock may be designated as Performance Tested Parent Stock, but this interrelationship shall be specified when the classification of either stock is referred to in advertising or certification.

(4) When products are sold or offered for sale as Performance Tested Parent Stock, the breeder shall be able to substantiate by records filed with the Official State Agency that such products are qualified for this classification.

(b) *U.S. Certified for Meat.* All males and females from Performance Tested Parent Stock for meat production mated in the same combination as used in the qualifying parent flock.

(c) *U.S. Approved.* All males and females selected by Authorized Agents or State Inspectors according to standards prescribed by the Official State Agency or the State College of Agriculture.

(d) *U.S. Pullorum-Typhoid Clean.* A flock in which freedom from pullorum and typhoid has been demonstrated by one of the following criteria (See § 145.14 relating to the official blood test.)

(1) It has been officially blood tested within the past 12 months with no reactors.

(2) It is a multiplier breeding flock meeting the following specifications:

(i) The flock is located in a State where all persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is isolated;

(ii) The flock is composed entirely of birds that originated from U.S. Pullorum-Typhoid Clean primary breeding flocks or from flocks that met equivalent requirements under official supervision; and

(iii) A sample comprised of at least 25 percent of the birds in the flock has been officially blood tested within the past 12 months with no reactors, or its progeny has been subjected to an approved 10-day chick mortality bacteriological examination monitoring program and bacteriological examination of a fluff sample from selected hatches as prescribed by the Official State Agency: *Provided,* That when the blood testing procedure is used, the percentage of the flock included in the sample may be reduced by 5 percentage points following each year in which there is no evidence of infection on the premises: *And provided further,* That the sample tested for the qualification of a flock under this subparagraph shall include at least 500 birds the first year, 400 the second year, 300 the third year, 200 the fourth year, and 100 the fifth year.

(3) It is a multiplier breeding flock composed entirely of birds that originated from U.S. Pullorum-Typhoid Clean primary breeding flocks or from flocks that met equivalent requirements under official supervision and is located in a State in which it has been determined by the ASR Division that:

(i) All hatcheries, except turkey hatcheries, within the State are qualified as "National Plan Hatcheries" or have

met equivalent requirements for pullorum-typhoid control under official supervision;

(ii) All hatchery supply flocks, except turkey flocks, within the State, are qualified as U.S. Pullorum-Typhoid Clean or have met equivalent requirements for pullorum-typhoid control under official supervision: *Provided*, That if other domesticated fowl are maintained on the same premises as the participating flock, freedom from pullorum-typhoid infection shall be demonstrated by an official blood test of each of these fowl;

(iii) All shipments of products other than U.S. Pullorum-Typhoid Clean, or equivalent, into the State are prohibited;

(iv) All persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is located;

(v) All reports of S. pullorum or S. gallinarum isolations from poultry are promptly followed by investigation by the Official State Agency to determine the origin of the infection;

(vi) All flocks found to be infected with pullorum or typhoid are quarantined until marketed or destroyed under the supervision of the Official State Agency, or until subsequently blood tested following the procedure for reacting flocks as contained in § 145.14(a)(5), and all birds fail to demonstrate pullorum or typhoid infection;

(vii) All poultry, except turkeys, going to public exhibition come from U.S. Pullorum-Typhoid Clean or equivalent flocks, or have had a negative pullorum-typhoid test within 90 days of going to public exhibition;

(viii) Discontinuation of any of the conditions or procedures described in subdivisions (i), (ii), (iii), (iv), (v), (vi), and (vii) of this subparagraph, or the occurrence of repeated outbreaks of pullorum or typhoid in poultry breeding flocks, other than turkey flocks, within or originating within the State shall be grounds for revocation of the determination. Such action shall not be taken until a thorough investigation has been made by the ASR Division and the Official State Agency has been given an opportunity for a hearing.

(4) It is a multiplier breeding flock located in a State which has been determined to be in compliance with the provisions of § 145.33(d)(3) and in which pullorum disease or fowl typhoid is not known to exist nor to have existed in hatchery supply flocks, other than turkey flocks, within the State during the preceding 24 months.

(5) It is a primary breeding flock located in a State determined to be in compliance with the provision of § 145.33(d)(4), and in which a sample of 300 birds from flocks of more than 300, and each bird in flocks of 300 or less, has been officially tested for pullorum-typhoid within the past 12 months with no reactors: *Provided*, That a bacteriological examination monitoring program acceptable to the Official State Agency and approved by the ASR Division may be used in lieu of blood testing.

(e) *U.S. M. Gallisepticum Clean.* (1) A flock maintained in compliance with the provisions of § 147.26 of this chapter and in which freedom from M. gallisepticum has been demonstrated by one of the following procedures:

(i) All birds have been tested as provided in § 145.14(b) when more than 5 months of age: *Provided*, That to retain this classification, a random sample of at least 10 percent of the flock shall be tested at intervals of not more than 60 days; or

(ii) It is a multiplier breeding flock which originated as U.S. M. Gallisepticum Clean chicks from primary breeding flocks, and samples comprising of at least 10 percent of the birds in the flock have been tested, as provided in § 145.14(b), twice between the ages of 8 weeks and 22 weeks, with an interval of not less than 60 days between the two tests. *Provided*, That to retain this classification, the flock shall be subjected to one of the following procedures:

(*a*) At intervals of not more than 90 days, a random sample of at least 5 percent of the flock shall be tested; or

(*b*) At intervals of not more than 30 days, a sample of 25 cull chicks produced from the flock shall be subjected to approved laboratory procedures for the detection and recovery of M. gallisepticum; or

(*c*) At intervals of not more than 60 days, serum samples obtained from at least 100-day-old chick produced from the flock shall be examined for M. gallisepticum antibodies.

(2) A participant handling U.S. M. Gallisepticum Clean products shall keep these products separate from other products in a manner satisfactory to be Official State Agency: *Provided*, That U.S. M. Gallisepticum Clean chicks from primary breeding flocks shall be produced in incubators and hatchers in which only eggs from flocks qualified under subdivision (i) of subparagraph (1) of this paragraph are set.

(3) U.S. M. Gallisepticum Clean chicks shall be boxed in clean boxes and delivered in clean, disinfected trucks.

Subpart D—Special Provisions for Turkey Breeding Flocks and Products

§ 145.41 Definitions.

Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:

(a) *Baby poults.* Poults that have not been fed or watered.

(b) *Broad-breasted.* A term used to describe a type of turkey which, at the time of selection and no later than 30 weeks of age, has a breast width at a point 1¾ inches above the keel of at least 3½ inches, for both toms and hens.

§ 145.42 Participation.

Participating turkey flocks, and the eggs and poults produced from them, shall comply with the applicable general provisions of Subpart A of this part and the special provisions of this Subpart D.

(a) All flocks shall consist of birds that have been selected for health, vigor, and freedom from physical deformities of economic importance by an Authorized Agent or State Inspector.

(b) The minimum weight of turkey hatching eggs shipped interstate shall be 2 ounces each for small varieties and 2½ ounces each for other varieties, unless otherwise specified by the purchaser of the eggs.

(c) All eggs set should have been fumigated as described in § 147.25(a) of this chapter, and should be refumigated as described in § 147.25(c) of this chapter or they should be fumigated as soon as possible (preferably within 12 hours) after setting as described in § 147.25(b) of this chapter. All eggs should be refumigated after transfer to the hatcher as described in § 147.25(d) of this chapter.

§ 145.43 Terminology and classification.

Participating flocks, and the eggs and poults produced from them, which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in § 145.10.

(a) *U.S. Approved.* All males and females selected by Authorized Agents or State Inspectors according to standards prescribed by the Official State Agency or the State College of Agriculture.

(b) *U.S. Pullorum-Typhoid Clean.* A flock in which freedom from pullorum and typhoid has been demonstrated by one of the following criteria (See § 145.14 relating to the official blood test.):

(1) It has been officially blood tested within the past 12 months with no reactors.

(2) It is a multiplier breeding flock meeting the following specifications:

(i) The flock is located in a State where all persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarium is isolated;

(ii) The flock is composed entirely of birds that originated from flocks that qualified as U.S. Pullorum-Typhoid Clean primary breeding flocks or from flocks that met equivalent blood testing requirements under official supervision; and

(iii) A sample comprised of at least 25 percent of the birds in the flock has been officially blood tested within the past 12 months with no reactors: *Provided*, That the percentage of the flock included in the sample may be reduced by 5 percentage points following each year in which there is no evidence of infection on the premises: *And provided further*, That the sample tested for the qualification of a flock under this subparagraph shall include at least 500 birds the first year, 400 the second year, 300 the third year, 200 the fourth year, 100 the fifth year; or

(3) It is a multiplier breeding flock composed entirely of birds that originated from flocks qualified as U.S. Pullorum-Typhoid Clean primary breeding flocks or from flocks that met equivalent blood testing requirements under official supervision, and is located in a State in which it has been determined by the ASR Division that:

(i) All chicken and turkey hatcheries within the State are qualified as "National Plan Hatcheries" or have met equivalent requirements for pullorum-typhoid control under official supervision;

(ii) All chicken and turkey hatchery supply flocks within the State are qualified as U.S. Pullorum-Typhoid Clean or have met equivalent requirements for blood testing under official supervision;

(iii) All shipments of products other than U.S. Pullorum-Typhoid Clean, or equivalent, into the State are prohibited;

(iv) All persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is isolated;

(v) All reports of S. pullorum or S. gallinarum isolations are promptly followed by an Official State Agency investigation to determine the origin of the infection;

(vi) All flocks found to be infected with pullorum or typhoid are quarantined until marketed or destroyed under the supervision of the Official State Agency, or subsequently blood tested and all birds in such flocks fail to demonstrate pullorum or typhoid infection;

(vii) All chickens and turkeys going to public exhibition come from U.S. Pullorum-Typhoid Clean or equivalent flocks, or have had a negative pullorum-typhoid test within 90 days of going to public exhibition; and

(viii) A monitoring program, including official blood tests of at least 25 percent of the birds in the hatchery supply flocks in the State, is systematically conducted each year. The samples tested are selected to be representative of all hatchery supply flocks in the State. The minimum requirements as to the percentage of birds tested in the monitoring program may be reduced by 5 percent of the total number of birds in all flocks following each year in which no infected birds are detected.

(c) *U.S. M. Gallisepticum Clean.* (1) A flock maintained in accordance with the conditions and procedures described in § 147.26 of this chapter, and in which no reactors are found when a random sample of at least 10 percent of the birds in the flock is tested when more than 4 months of age, in accordance with the procedures described in § 145.14(b).

(2) A flock qualified as U.S. M. Gallisepticum Clean may retain the classification for 1 year, provided it is maintained in isolation and no evidence of M. gallisepticum infection is revealed. Each flock and premises shall be inspected at least once during the laying period by an Authorized Agent of the Official State Agency or the State Animal Disease Control Official. If a flock proves to be infected with M. gallisepticum, it shall be eliminated as a breeding flock under the supervision of the Official State Agency or the State Animal Disease Control Official.

(3) In order to sell hatching eggs or poults of this classification, all hatching eggs and poults handled by the participant must be of this classification.

(d) *U.S. Typhimurium Controlled.* (1) A flock meeting the following requirements:

(i) (*a*) All birds have been officially blood tested within 12 months for S. typhimurium as provided in § 145.14(a) (2) and no reactors were found on the first test; or

(*b*) All birds are located on premises and originated from premises where U.S. Typhimurium Controlled flocks, tested in accordance with § 145.43(d) (1) (i) (*a*), were maintained for a 2-year period with no evidence of S. typhimurim infection, and no subsequent evidence of infection found. Flocks must be located within a State in which all isolations of S. typhimurium are reported promptly to both the Official State Agency and the State Animal Disease Control Official.

(ii) The flock is maintained in compliance with the provisions of § 147.21 of this chapter, and the hatching eggs are handled in compliance with the provisions of § 147.22 of this chapter in a manner satisfactory to the Official State Agency. All eggs used for hatching shall be visibly clean and fumigated as described in § 147.25(a) of this chapter as soon as possible after collection. Each flock and premises shall be inspected at least once during the egg production season by a State Inspector to ascertain that these provisions are being followed. The Official State Agency shall immediately terminate the U.S. Typhimurium Controlled classification of flocks found to be in noncompliance with these provisions.

(2) In order to sell hatching eggs or poults of this classification, all hatching eggs and poults handled must meet the requirements for this classification.

(3) Hatcheries producing products of this classification shall be maintained in compliance with the provisions of §§ 147.23, 147.24, and 147.25 of this chapter in a manner satisfactory to the Official State Agency.

Subpart E—Special Provisions for Waterfowl, Exhibition Poultry, and Game Bird Breeding Flocks and Products

§ 145.51 Definitions.

Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:

(a) *Waterfowl.* Domesticated fowl that normally swim, such as ducks and geese.

(b) *Exhibition poultry.* Domesticated fowl which are bred for the combined purposes of meat or egg production and competitive showing.

(c) *Game birds.* Domesticated fowl such as pheasants, partridge, quail, grouse, and guineas, but not doves and pigeons.

§ 145.52 Participation.

Participating flocks of waterfowl, exhibition poultry, and game birds, and the eggs and baby poultry produced from them, shall comply with the applicable general provisions of Subpart A of this part and the special provisions of this Subpart E.

(a) All eggs should be fumigated at the hatchery as described in § 147.25(a) or (b) or (c) of this chapter. All eggs should be refumigated after transfer to the hatchery as described in § 147.25(d) of this chapter.

(b) Started poultry shall lose their identity under Plan terminology when not maintained by Plan participants under the conditions prescribed in § 145.5(a).

§ 145.53 Terminology and classification.

Participating flocks, and the eggs and baby poultry produced from them, which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in § 145.10.

(a) *U.S. Approved.* All males and females selected by Authorized Agents or State Inspectors according to standards prescribed by the Official State Agency or the State College of Agriculture.

(b) *U.S. Pullorum-Typhoid Clean.* A flock in which freedom from pullorum and typhoid has been demonstrated by one of the following criteria (See § 145.14 relating to the official blood test.):

(1) It has been officially blood tested within the past 12 months with no reactors.

(2) It is a multiplier breeding flock meeting the following specifications:

(i) The flock is located in a State where all persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is isolated;

(ii) The flock is composed entirely of birds that originated from U.S. Pullorum-Typhoid Clean primary breeding flocks or from flocks that met equivalent requirements under official supervision; and

(iii) A sample comprised of at least 25 percent of the birds in the flock has been officially blood tested within the past 12 months with no reactors, or its progeny has been been subjected to an approved 10-day poultry mortality bacteriological examination monitoring program and bacteriological examination of a fluff sample from selected hatches as prescribed by the Official State Agency: *Provided,* That when the blood testing procedure is used, the percentage of the flock included in the sample may be reduced by 5 percentage points following each year in which there is no evidence of infection on the premises: *And provided further,* That the sample tested for

the qualification of a flock under this subparagraph shall include at least 500 birds the first year, 400 the second year, 300 the third year, 200 the fourth year, and 100 the fifth year.

(3) It is a multiplier breeding flock composed entirely of birds that originated from U.S. Pullorum-Typhoid Clean primary breeding flocks or from flocks that met equivalent requirements under official supervision, and is located in a State in which it has been determined by the ASR Division that:

(i) All hatcheries, except turkey hatcheries, within the State are qualified as "National Plan Hatcheries" or have met equivalent requirements for pullorum-typhoid control under official supervision;

(ii) All hatchery supply flocks, except turkey flocks, within the State, are qualified as U.S. Pullorum-Typhoid Clean or have met equivalent requirements for pullorum-typhoid control under official supervision: *Provided,* That if other domesticated fowl are maintained on the same premises as the participating flock, freedom from pullorum-typhoid infection shall be demonstrated by an official blood test of each of these fowl;

(iii) All shipments of products other than U.S. Pullorum-Typhoid Clean, or equivalent, into the State are prohibited;

(iv) All persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is isolated;

(v) All reports of S. pullorum or S. gallinarum isolations from poultry are promptly followed by an investigation by the Official State Agency to determine the origin of the infection;

(vi) All flocks found to be infected with pullorum or typhoid are quarantined until marketed or destroyed under the supervision of the Official State Agency, or until subsequently blood tested, following the procedure for reacting flocks as contained in § 145.14(a)(5), and all birds fail to demonstrate pullorum or typhoid infection;

(vii) All poultry, except turkeys, going to public exhibition come from U.S. Pullorum-Typhoid Clean or equivalent flocks, or have had a negative pullorum-typhoid test within 90 days of going to public exhibition;

(viii) Discontinuation of any of the conditions or procedures described in subdivisions (i), (ii), (iii), (iv), (v), (vi), and (vii) of this subparagraph, or the occurrence of repeated outbreaks of pullorum or typhoid in poultry breeding flocks, other than turkey flocks, within or originating within the State shall be grounds for revocation of the determination. Such action shall not be taken until a thorough investigation has been made by the ASR Division and the Official State Agency has been given an opportunity for a hearing.

(4) It is a multiplier breeding flock located in a State which has been determined to be in compliance with the provisions of § 145.53(b)(3), and in which pullorum disease or fowl typhoid is not known to exist nor to have existed in hatchery supply flocks, other than turkey flocks, within the State during the preceding 24 months.

(5) It is a primary breeding flock located in a State determined to be in compliance with the provision of § 145.53(b)(4), and in which a sample of 300 birds from flocks of more than 300, and each bird in flocks of 300 or less, has been officially tested for pullorum-typhoid within the past 12 months with no reactors: *Provided,* That a bacteriological examination monitoring program acceptable to the Official State Agency and approved by the ASR Division may be used in lieu of blood testing.

(c) *U.S. M. Gallisepticum Clean.* (1) A flock maintained in compliance with the provisions of § 147.26 of this chapter and in which freedom from M. gallisepticum has been demonstrated by one of the following procedures:

(i) All birds have been tested as provided in § 145.14(b) when more than 5 months of age: *Provided,* That to retain this classification, a random sample of at least 10 percent of the flock shall be tested at intervals of not more than 60 days; or

(ii) It is a multiplier breeding flock which originated as U.S. M. Gallisepticum Clean baby poultry from primary breeding flocks, and samples comprising at least 10 percent of the birds in the flock have been tested, as provided in § 145.14(b), twice between the ages of 8 weeks and 22 weeks, with an interval of not less than 60 days between the two tests: *Provided,* That to retain this classification, the flock shall be subjected to one of the following procedures:

(*a*) At intervals of not more than 90 days, a random sample of at least 5 percent of the flock shall be tested; or

(*b*) At intervals of not more than 30 days, a sample of at least 25 cull baby poultry produced from the flock shall be subjected to approved laboratory procedures for the detection and recovery of M. gallisepticum; or

(*c*) At intervals of not more than 60 days, serum samples obtained from at least 100-day-old baby poultry produced from the flock shall be examined for M. gallisepticum antibodies.

(2) A participant handling U.S. M. Gallisepticum Clean products shall keep these products separate from other products in a manner satisfactory to the Official State Agency: *Provided,* That U.S. M. Gallisepticum Clean baby poultry from primary breeding flocks shall be produced in incubators and hatchers in which only eggs from flocks qualified under subdivision (i) of subparagraph (1) of this paragraph are set.

(3) U.S. M. Gallisepticum Clean baby poultry shall be boxed in clean boxes and delivered in clean, disinfected trucks.

PART 147—AUXILIARY PROVISIONS OF THE NATIONAL POULTRY IMPROVEMENT PLAN

Subpart A—Blood Testing Procedures

Sec.
147.1 The standard tube agglutination test.
147.2 The rapid serum test.
147.3 The stained-antigen, rapid, whole-blood test.
147.4 The tube agglutination test for S. typhimurium.

Subpart B—Bacteriological Examination Procedure

147.11 Laboratory procedure recommended for the bacteriological examination of reactors.

Subpart C—Sanitation Procedures

147.21 Flock sanitation.
147.22 Hatching egg sanitation.
147.23 Hatchery sanitation
147.24 Cleaning and disinfecting.
147.25 Fumigation.
147.26 Procedures for establishing isolation and maintaining sanitation and good management practices for the control of Mycoplasm gallisepticum.
147.27 Procedures recommended to prevent the spread of disease by artificial insemination of turkeys.

Subpart D—Random Sample Performance Testing Procedures

147.31 Random sample tests; general.
147.32 Random sample egg production test.
147.33 Random sample meat production test.
147.34 Random sample tests; combined summary.

Subpart E—Procedure for Changing National Poultry Improvement Plan

147.41 Definitions.
147.42 General.
147.43 General Conference Committee.
147.44 Submitting, compiling, and distributing proposed changes.
147.45 Official delegates.
147.46 Committee consideration of proposed changes.
147.47 Conference consideration of proposed changes.
147.48 Approval of conference recommendations by the Department.

Subpart A—Blood Testing Procedures

§ 147.1 The standard tube agglutination test.[1]

(a) The blood samples should be collected and delivered as follows:

(1) The blood samples should be taken by properly qualified and authorized persons only, and in containers provided by the laboratory. The containers should be stoutwalled test tubes, preferably $\frac{3}{8}$ inch by 3 inches, without lip, or small well-selected medicine vials, which have been throughly cleaned and dried in a hot-air drying oven. If stoppers are used, they should be thoroughly cleaned and dried.

(2) Sufficient blood should be procured by making a small incision in the large median wing vein with a small sharp lancet and allowing the blood to run into

[1] The procedure described is a modification of the method reported in the Proceedings of the United States Live Stock Sanitary Association, Nov. 30 to Dec. 2, 1932, pp. 487 to 491.

the tube, or by the use of a small syringe (with 20- or 21-gage needle) which is properly cleansed between bleedings with physiological saline solution. To facilitate the separation of the serum, the tubes should be placed in a slanted position until the blood has solidified. After the blood has completely clotted, they should be packed and shipped by mail (special delivery), rapid express, or by messenger, to the laboratory. All labeling must be clear and permanent, and may be done with a suitable pencil on etched portions of the tube, or by means of fast-gum labels.

(3) The blood samples must reach the laboratory in a fresh and unhemolyzed condition. Hemolyzed samples should be rejected. It is imperative, therefore, to cool the tubes immediately after slanting and clotting, and unless they reach the laboratory within a few hours, to pack them with ice in special containers, or use some other cooling system which will insure their preservation during transportation. In severe cold seasons, extreme precautions must be exercised to prevent freezing and consequent laking. The samples must be placed in cold (5° to 10° C.) storage, immediately upon arrival at the laboratory.

(b) The antigen shall consist of representative strains of S. pullorum which are of known antigenic composition, high agglutinability, but are not sensitive to negative and nonspecific sera. The stock cultures may be maintained satisfactorily by transferring to new sloped agar at least once a month and keeping at 18° to 25° C. (average room temperature) in a dark closet or chest, following incubation for from 24 to 36 hours at 37° C. The antigenic composition and purity of the stock cultures should be checked consistently.

(c) A medium which has been used satisfactorily has the following composition:

Water	1,000 cc.
Difco beef extract	4 gm. (0.4 percent).
Difco Bacto-peptone	10 gm. (1.0 percent).
Difco dry-granular agar.	20 gm. (2.0 percent).
Reaction—pH 6.8 to 7.2.	

(d) Large 1-inch test tubes, Kolle flasks, or Blake bottles should be streaked liberally over the entire agar surface with inoculum from 48-hour slant agar cultures prepared from the stock cultures of the selected strains. The antigen-growing tubes or bottles should be incubated 48 hours at 37° C., and the surface growth washed off with sufficient phenolized (0.5 percent) saline (0.85 percent) solution to make a heavy suspension. The suspension should be filtered free of clumps through a thin layer of absorbent cotton in a Buchner funnel with the aid of suction. The antigens of the separate strains should be combined in equal volume-density and stored in the refrigerator (5° to 10° C.) in tightly stoppered bottles.

(e) Thiosulfate-Glycerin (TG) medium may be used as an alternate medium for the preparation of tube agglutination antigen. The TG medium, formerly used for the preparation of stained, whole-blood antigen, is described in more detail in the article by MacDonald, A.D., Recent Developments in Pullorum Antigen for the Rapid, Whole-Blood Test, Report of the Conference of the National Poultry Improvement Plan, pages 122–127, 1941. This medium provides a tube antigen of excellent specificity and greatly increases the yield of antigen from a given amount of medium. The TG medium has the following composition:

Beef infusion	1,000 cc.
Difco Bacto-peptone	20 gm. (2.0 percent).
Sodium thiosulfate	5 gm. (0.5 percent).
Ammonium chloride	5 gm. (0.5 percent).
Glycerin, U.S.P. (95 percent)	20 cc. (2.0 percent).
Difco dry-granular agar	30 gm. (3.0 percent).
Reaction—pH 6.8 to 7.2.	

Large 1-inch test tubes, Kolle flasks, Blake bottles, or Erlenmeyer flasks should be seeded over the entire agar surface with inoculum from 24-hour beef infusion broth cultures prepared from the stock cultures of the selected strains. The antigen-growing tubes or bottles should be incubated 96 hours at 37° C., and the surface growth washed off with sufficient phenolized (0.5 percent) saline (0.85 percent) solution to make a heavy suspension. The suspension should be filtered free of clumps through a thin layer of absorbent cotton in a Buchner funnel with the aid of suction. The antigen then should be centrifuged. The mass of bacteria should be removed from the centrifuge tubes or bowl and resuspended in saline (0.85 percent) solution containing 0.5 percent phenol. After the bacterial mass has been uniformly suspended in the diluent, it should be again passed through a cotton pad in a Buchner funnel without the aid of suction. The antigens of the separate strains should be combined in equal volume-density and stored in the refrigerator (5° to 10° C.) in tightly stoppered bottles.

(f) The diluted antigen to be used in the routine testing should be prepared from the stock antigen by dilution of the latter with physiological (0.85 percent) saline solution containing 0.25 percent of phenol to a turbidity corresponding to 0.75–1.00 on the McFarland nephelometer scale. The hydrogen-ion concentration of the diluted antigen should be corrected to pH 8.2 to 8.5 by the addition of dilute sodium hydroxide. New diluted antigen should be prepared each day and kept cold. The diluted anitgen may be employed in 2 cc. quantities in 4- by ½-inch test tubes, or 1 cc. quantities in smaller tubes, in which the final serum-antigen mixtures are made and incubated. The distribution of the antigen in the tubes may be accomplished by the use of long burettes, or special filling devices made for the purpose.

(g) The maximum serum dilution employed must not exceed 1:50 for chickens, nor 1:25 for turkeys. The available data indicate that 1:25 dilution is the most efficient. In all official reports on the blood test, the serum dilutions shall be indicated. The sera should be introduced into the agglutination tubes in the desired amounts with well-cleaned serological pipettes or special serum-delivery devices which do not permit the mixing of different sera. The antigen and serum should be well mixed before incubation. The serum and antigen mixture must be incubated for at least 20 hours at 37° C.

(h) The results shall be recorded as:

N, or — (negative) when the serum-antigen mixture remains uniformly turbid.

P, or + (positive) when there is a distinct clumping of the antigen, and the liquid between the agglutinated particles is clear.

S, or ? (suspicious) when the agglutination is only partial or incomplete.

M, or missing, when samples listed on the original record sheet are missing.

H, or hemolyzed, when blood samples are hemolyzed and cannot be tested.

B, or broken, when sample tubes are broken and no serum can be obtained.

(Some allowances must always be made for the difference in sensitiveness of different antigens and different setups, and therefore, a certain amount of independent, intelligent judgment must be exercised at all times. Also, the histories of the flocks require consideration. In flocks where individuals show a suspicious agglutination, it is desirable to examine representative birds bacteriologically to determine the presence or absence of S. pullorum.)

§ 147.2 The rapid serum test.[2]

(a) The procedure for the collection and delivery of blood samples in the rapid serum test is the same as that described in § 147.1(a).

(b) The selection and maintenance of suitable strains of S. pullorum and the composition of a satisfactory medium are described in § 147.1 (b) and (c).

(c) Large 1-inch test tubes, Kolle flasks, or Blake bottles are streaked liberally from 48-hour slant-agar cultures prepared from stock cultures of the selected strains.

(d) The antigen-growing tubes or bottles should be incubated 48 hours at 37° C., and the surface growth washed off with a very slight amount of 12 percent solution of sodium chloride containing 0.25 to 0.5 percent phenol, filtered through lightly packed sterile absorbent cotton placed in the apex of a sterile funnel.

(e) The washings should be adjusted (using 12 percent sodium chloride containing 0.25 to 0.5 percent phenol) so that the turbidity is 50 times greater than tube 0.75 of McFarland's nephelometer, or to a reading of 7 mm. by the Gates nephelometer.

(f) The individual strain antigens should be tested with negative sera for their insensitivity and with positive sera for high agglutinability in comparison with known satisfactory antigen. The antigens of the separate strains should be combined in equal volume-density

[2] The procedure described is a modification of the method reported by Runnels, Coon, Farley, and Thorpe, Amer. Vet. Med. Assoc. Jour. 70 (N.S. 23): 660–662 (1927).

and stored in the refrigerator (5° to 10° C.) in tightly stoppered bottles.

(g) The tests should be conducted on a suitable, smooth plate. The serum-antigen dilution should be made so that the dilution will not exceed 1:50 when compared to the standard tube agglutination test. When testing turkey blood samples, it is desirable to use a serum-antigen dilution equivalent to the 1:25 in the tube method. The serum should be added to the antigen and mixed thoroughly by use of the tip of the serum pipette. Most strong positive reactions will be plainly evident within 15 to 20 seconds. The final reading should be made at the end of 2 or 3 minutes. Heating the plate at approximately 37° C. will hasten agglutination. Before reading, the plate should be rotated several times.

(h) The results shall be recorded in § 147.1(h).

§ 147.3 The stained-antigen, rapid, whole-blood test.[3]

(a) The description of the preparation of antigen is not herein included because the antigen is a proprietary product produced only under license from the Secretary of Agriculture.

(b) A loop for measuring the correct quantity of blood can usually be obtained from the manufacturer of the antigen. A satisfactory loop may be made from a piece of No. 20 gage nichrome wire, 2½ inches long, at the end of which is fashioned a loop three-sixteenths of an inch in diameter. Such a loop, when filled with blood so that the blood appears to bulge, delivers 0.02 cc. A medicine dropper whose tip is adjusted to deliver 0.05 cc. is used to measure the antigen. A glass plate about 15-inches square, providing space for 48 tests, has proved satisfactory for this work. The use of such a plate enables the tester to have a number of successive test mixtures under observation without holding up the work to wait for results before proceeding to the next bird.

(c) A drop of antigen should be placed on the testing plate. A loopful of blood should be taken up from the wing vein. When submerged in the blood and then carefully withdrawn, the loop becomes properly filled. On looking down edgewise at the filled loop, one observes that the blood appears to bulge. The loopful of blood then should be stirred into the drop of antigen, and the mixture spread to a diameter of about 1 inch. The loop then should be rinsed in clean water and dried by touching it to a piece of clean blotting paper, if necessary. The test plate should be rocked from side to side a few times to mix the antigen and blood thoroughly, and to facilitate agglutination. The antigen should be used according to the directions of the producer.

(d) Various degrees of reaction are observed in this as in other agglutination tests. The greater the agglutinating ability of the blood, the more rapid the clumping and the larger the clumps. A positive reaction consists of a definite clumping of the antigen surrounded by clear spaces. Such reaction is easily distinguished against a white background. A somewhat weaker reaction consists of small but still clearly visible clumps of antigen surrounded by spaces only partially clear. Between this point and a negative or homogeneous smear, there sometimes occurs a very fine granulation barely visible to the naked eye; this should be disregarded in making a diagnosis. The very fine marginal clumping which may occur just before drying up is also regarded as negative. In a nonreactor, the smear remains homogeneous. (Allowance should be made for differences in the sensitiveness of different antigens and different setups, and therefore, a certain amount of independent, intelligent judgment must be exercised at all times. Also, the histories of the flocks require consideration. In flocks where individuals show a suspicious agglutination, it is desirable to examine representative birds bacteriologically to determine the presence or absence of S. pullorum.)

§ 147.4 The tube agglutination test for S. typhimurium.

(a) The procedure for the collection and delivery of blood samples in the tube agglutination test for S. typhimurium is the same as that described in § 147.1(a).

(b) The "O" antigen should be prepared as follows:

(1) The antigen shall consist of a representative nonmotile strain of S. typhimurium which is of known antigenic composition and high agglutinability but is not sensitive to negative and nonspecific sera. Strain P 10 meets these requirements.

(2) The stock culture is maintained on 1 percent nutrient agar deeps, which have been incubated for 18–24 hours at 37° C. They are stored at room temperature.

(3) A satisfactory medium used for growing the organism is veal infusion agar (Difco). It is dispensed in 50 ml. amounts into 500 ml. medicine bottles, with screw caps, and sterilized at 15 pounds pressure for 20 minutes. The bottles are then laid flat upon an even surface until the medium has solidified.

(4) The inoculum used for preparation of "O" antigen is a nonmotile strain of S. Typhimurium. The organism is grown in veal infusion broth (Difco) for 18–24 hours at 37° C.; then plated, for single colony isolation, on veal infusion agar plates. These plates are incubated for 18–24 hours at 37° C. After incubation, single colonies are picked and transferred to veal infusion agar slants, which are incubated for 18–24 hours at 37° C. After this, the cultures are tested for smoothness by using a 1:500 dilution of acriflavine.

(5) Smooth cultures are inoculated into flasks containing veal or beef infusion broth which is incubated for 18–24 hours at 37° C. The incubated broth suspension of organisms is dispensed into the antigen bottles containing veal infusion agar. The suspension is distributed evenly over the agar surface by gently tilting the bottles from side to side. The inoculated bottles are then laid flat, agar side down, for 10–20 minutes. They are subsequently incubated, agar side upward, for 24–48 hours at 37° C. before harvesting.

(6) The harvesting of the organism consists of washing the growth from each antigen bottle with 0.5 percent phenolized physiological saline. The bacterial suspension from each bottle is filtered through sterile milk pad filters into a large sterile container or through a thin layer of absorbent cotton in a Buchner funnel with the aid of suction. To each 100 ml. of the bacterial suspension is added additional phenol to make the final concentration 0.5 percent. The concentrated antigen is tested for sterility at intervals after 24 hours. After sterility is proved, the stock antigen is standardized to determine the density according to the McFarland nephelometer scale.

(7) The diluted antigen to be used in routine testing is prepared from stock antigen, by diluting with 0.25 percent phenolized saline, and is standardized to a turbidity corresponding to 0.75–1.00 of the McFarland nephelometer scale.

(c) The maximum serum dilution employed for the "O" antigen tube test must not exceed 1:25. In all official reports on the blood test, the serum dilutions should be indicated. The sera should be introduced into the agglutination tubes in the desired amounts with well-cleaned serological pipettes or special serum delivery devices which do not permit the mixing of different sera. The antigen and serum should be well mixed before incubation. The serum and antigen mixture must be incubated for at least 20 hours at 37° C.

(d) The results shall be recorded as described in § 147.1(h).

Subpart B—Bacteriological Examination Procedure

§ 147.11 Laboratory procedure recommended for the bacteriological examination of reactors.

(a) The pericardial sac, peritoneum, oviduct, and any visibly pathological tissues should be cultured on beef extract agar or tryptose agar by means of sterile swabs. Sterile technique should be followed. (Primary culture of these organs in a suitable nutrient broth and transfer to a suitable nutrient agar is optional.)

(b) The following organs should be aseptically collected for culture:

(1) Heart (apex, pericardial sac, and contents if present);

(2) Liver (portions exhibiting lesions or, in grossly normal organs, the drained gall bladder and adjacent liver tissues);

(3) Ovary-Testes (entire inactive ovary or testes, but if ovary is active, use own judgment and include any atypical ova);

(4) Oviduct (if active, include any debris and dehydrated ova);

(5) Pancreas; and

(6) Spleen.

(c) A composite sample of the organs listed in § 147.11(b) should be ground in a sterile mortar or suitable blender. Individual organs may be used if desired. Nutrient broth should be added as a

[3] The procedure described is a modification of the method reported by Schaffer, MacDonald, Hall, and Bunyea, Jour. Amer. Vet. Med. Assoc. 79 (M.S. 32); 236–240 (1931).

diluent. Ten cc. of this suspension should be inoculated into 100 cc. of either Selenite F broth or Tetrathionate broth, and into 100 cc. of a suitable noninhibitory nutrient broth.

(d) After 24 hours incubation at 37° C., a loopful of the broth cultures from each flask should be streaked on a suitable noninhibitory solid medium, such as tryptose agar, and one of the following selective media: Salmonella-Shigella (SS), MacConkey, Brilliant Green, Bismuth Sulfite, or Desoxycholate Citrate Lactose Sucrose (D.C.L.S.) agar. (All of these media may be obtained in dehydrated form.) If no suspicious colonies are observed after 24 hours incubation, the enrichment broths should be restreaked on solid media.

(e) A portion of the crop wall and intestine to include the cecal tonsils are put into either Selenite F or Tetrathionate broth and incubated for 24 hours at 37° C. Transfers should be made from the broth onto agar plates as indicated in § 147.11(d).

(f) Suspicious single colonies should be subcultured on nutrient agar or triple sugar iron agar slants and incubated for 24 hours at 37° C.

(g) Cultures should be transferred to the following fermentable media for identification: dextrose, lactose, sucrose (saccharose), mannite (mannitol), maltese, dulcite (dulcitol), and salicin broths. Suitable tests also should be conducted for the detection of indole, hydrogen sulfide, acetylmethylcarbinol, and urease production. Motility or nonmotility is demonstrated by inoculation of a suitable semisolid medium. For the Gram stain, a 24-hour nutrient agar slant culture should be used.

(h) All Salmonella cultures isolated should be serologically typed.

Subpart C—Sanitation Procedures

§ 147.21 Flock sanitation.

To aid in the maintenance of healthy flocks, the following procedures should be practiced:

(a) Baby poultry should be started in a clean brooder house and maintained in constant isolation from older birds and other animals. Personnel that are in contact with older birds and other animals should take precautions, including disinfection of footwear and change of outer clothing, to prevent the introduction of infection through droppings that may adhere to the shoes, clothing, or hands. (See § 147.24(a).)

(b) Range used for growing young stock should not have been used for poultry the preceding year. Where broods of different ages must be kept on the same farm, there should be complete depopulation of brooder houses and other premises following infection of such premises by any contagious disease.

(c) Poultry houses should be screened and proofed against free-flying birds. An active rodent eradication campaign is an essential part of the general sanitation program. The area adjacent to the poultry house should be kept free from accumulated manure, rubbish, and unnecessary equipment. Dogs, cats, sheep, cattle, horses, and swine should never have access to poultry operations. Visitors should not be admitted to poultry areas, and authorized personnel should take the necessary precautions to prevent the introduction of disease.

(d) Poultry houses and equipment should be thoroughly cleaned and disinfected prior to use for a new lot of birds. (See § 147.24(a).) Feed and water containers should be situated where they cannot be contaminated by droppings and should be frequently cleaned and disinfected. Dropping boards or pits should be constructed so birds do not have access to the droppings.

(e) Poultry house floors, other than slats or wire, should be well covered with an absorbent type of litter. Frequent stirring of the litter may be necessary to reduce excess moisture and prevent surface accumulation of droppings. Slat or wire floors should be constructed so as to permit free passage of droppings and to prevent the birds from coming in contact with the droppings. Nesting areas should be kept clean and, where appropriate, filled with clean nesting material.

(f) When an outbreak of disease occurs in a flock, dead or sick birds should be taken, by private carrier, to a diagnostic laboratory for complete examination. All Salmonella and Arizona cultures isolated should be typed serologically, and complete records maintained by the laboratory as to types recovered from each flock within an area. Records on isolations and serological types should be made available to Official State Agencies or other animal disease control regulatory agencies in the respective States for followup of foci of infection. Such information is necessary for the development of an effective Salmonella control program.

(g) Introduction of started or mature birds should be avoided to reduce the possible hazard of introducing infectious diseases. If birds are to be introduced, the health status of both the flock and introduced birds should be evaluated.

(h) In rearing broiler or replacement stock, a sound and adequate immunization program should be adopted. Since different geographic areas may require certain specific recommendations, the program recommended by the State experiment station or other State agencies should be followed.

§ 147.22 Hatching egg sanitation.

Hatching eggs should be collected from the nests at frequent intervals and, to aid in the prevention of contamination with disease causing organisms, the following practices should be observed:

(a) Cleaned and disinfected containers should be used in collecting the eggs, and precautions taken to prevent contamination from organisms that may be present on the hands or clothing of the person making the collection.

(b) Dirty eggs should not be used for hatching purposes and should be collected in a separate container from hatching eggs. Slightly soiled eggs may be dry cleaned by hand or motor driven buffer.

(c) The visibly clean eggs should be fumigated as described in § 147.25(a) as soon as possible after collection.

(d) The fumigated eggs should be stored in a cool place. Eggs should be stored no longer than necessary before setting. Packs used for storing eggs should be properly cleaned and disinfected.

(e) New or clean, fumigated cases should be used to transport eggs to the hatchery. Soiled egg case fillers should be destroyed.

§ 147.23 Hatchery sanitation.

An effective program for the prevention and control of Salmonella and other infections should include the following measures:

(a) The hatchery building should be arranged so that separate rooms, with separate ventilation, are provided for each of the four operations: Egg receiving, incubation and hatching, holding of baby poultry, and disposal of offal and cleaning of trays. These rooms should be placed under isolation so that admission is granted only to specifically authorized personnel who have taken proper precautions to prevent introduction of disease.

(b) The hatchery rooms, and tables, racks, and other equipment in them should be thoroughly cleaned and disinfected frequently. All hatchery wastes and offal should be burned or otherwise properly disposed of, and the containers used to remove such materials should be cleaned and sterilized after each use.

(c) The hatching compartments of incubators, including the hatching trays, should be thoroughly cleaned and fumigated after each hatch.

(d) Only clean eggs should be used for hatching purposes. All eggs set should be fumigated prior to setting or as soon as possible (preferably within 12 hours) after they are placed in the incubator. They should also be fumigated after transfer to a separate hatcher. (See § 147.25(d).)

(e) Only new or clean, fumigated egg cases should be used for transportation of hatching eggs. Soiled egg case fillers should be destroyed.

(f) Day-old chicks, poults, or other newly hatched poultry should be distributed in clean, new boxes. All crates and vehicles used for transporting started or adult birds should be cleaned and disinfected after each use.

§ 147.24 Cleaning and disinfecting.

(a) In the poultry houses and hatchery rooms:

(1) Settle dust by spraying lightly with the disinfectant to be used.

(2) Remove all litter and droppings to an isolated area where there is no opportunity for dissemination of any infectious disease organisms that may be present.

(3) Scrub the walls, floors, and equipment with a hot soapy water solution. Rinse to remove soap.

(4) Spray with a cresylic disinfectant, such as liquor cresolis saponatus,

4 ounces to the gallon of water, or sodium orthophenylphenate, 1⅓ ounces (1 heaping tablespoonful) to a gallon of hot water.

(b) In the hatchers:

(1) Remove trays and all controls and fans for separate cleaning. The ceiling, walls, and floors should be thoroughly wetted with a stream of water; then scrubbed with a hard bristle brush. Rinse until there is no longer any deposit on the walls, particularly near the fan opening.

(2) Replace the cleaned fans and controls. Replace the trays, preferably still wet from cleaning, and bring the incubator to normal operating temperature.

(3) The hatcher should be fumigated as described in § 147.25(e) prior to the transfer of the eggs.

(c) If the same machine is used for incubating and hatching, the entire machine should be cleaned after each hatch. A vacuum cleaner should be used to remove dust and down from the egg trays; then the entire machine should be vaccumed, mopped, and fumigated according to the procedures described in § 147.25(b) (3), (4), and (5).

§ 147.25 Fumigation.

Fumigation is recommended for sanitizing eggs and hatchery equipment as an essential part of a sanitation program.

(a) Fumigation of clean eggs after collection should be done as follows:

(1) Provide a room or cabinet proportionate to the number of eggs to be handled. The room should be relatively tight and must be equipped with a fan to circulate the gas during fumigation and to expel it after fumigation.

(2) The eggs should be placed on wire racks, in wire baskets, or on cup-type egg flats stacked outside of the egg cases (to permit air circulation), and exposed to circulating formaldehyde gas.

(3) Formaldehyde gas is provided by mixing 0.6 gram of potassium permanganate with 1.2 cc. of formalin (37.5 percent) for each cubic foot of space in the room. The ingredients should be mixed in an earthenware or enamelware container having a capacity at least ten times the volume of the total ingredients.

(4) Circulate the gas within the room for 20 minutes; then expel.

(5) The temperature in the cabinet during fumigation should be at least 70° F., and the relative humidity above 70 percent.

(b) Eggs should be fumigated at the hatchery prior to setting or as soon as possible after setting (preferably within 12 hours). Single or repeated fumigation of eggs in the setter may be practiced, but the fumigation schedule should be such that no eggs are fumigated during the period from the 24th to the 84th hour of incubation. The following procedure should be used:

(1) Determine the size of the incubator by multiplying the length times the width times the height.

(2) After setting the eggs and allowing temperature and humidity to regain normal operating levels, release formaldehyde gas into the incubator.

(3) For each cubic foot of space in the incubator, use 0.4 grams of potassium permanganate and 0.8 cc. of formalin (37.5 percent). For mixing the fumigant, use an earthenware or enamelware container having the capacity of at least 10 times the volume of the total ingredients.

(4) Close vents and doors but keep circulating fan operating, and continue fumigation for 20 minutes with normal operating temperature and humidity.

(5) After 20 minutes of fumigation, the vents should be opened to the normal operating positions to release the gas.

(c) Eggs which have not been fumigated in the hatchery as described in paragraph (b) of this section should be fumigated after the 84th hour of incubation. The procedure described in paragraph (b) of this section should be followed.

(d) All eggs should be fumigated after transfer to a separate hatcher, preferably as soon as the temperature and humidity regain normal operating levels. The procedure described in paragraph (b) of this section should be followed.

(e) Empty hatchers should be fumigated between each hatch. After the interior of the hatcher has been thoroughly cleaned and the cleaned trays returned, the following procedure should be followed:

(1) After temperature and humidity are brought to normal operating levels, use 0.6 grams of potassium permanganate and 1.2 cc. of formalin (37.5 percent) per cubic foot of space in the hatcher.

(2) Close the doors and vents and leave closed at least 3 hours, preferably overnight.

(f) The cheesecloth method of fumigation described in this paragraph may be used in lieu of the chemical method described in paragraph (b) of this section, using 0.6 cc. of formalin (37.5 percent) per cubic foot of space in the incubator, or paragraph (e) of this section, using 0.9 cc. of formalin (37.5 percent) for each cubic foot of space in the empty hatcher.

(1) Enough cheesecloth should be used to absorb all of the formalin that is to be used for the fumigation.

(2) The formalin-saturated cheesecloth should be hung in the cabinet in such a manner as to permit the circulating air to evaporate all the formalin. This will require longer than 20 minutes.

(3) Care should be taken to prevent the cheesecloth from blocking the air movement created by the fans.

(4) The cheesecloth method is not suitable for still air machines.

§ 147.26 Procedures for establishing isolation and maintaining sanitation and good management practices for the control of Mycoplasma gallisepticum.

(a) Required procedures:

(1) Allow no visitors except under controlled conditions;

(2) Maintain breeder flocks on farms free from market birds, or follow proper isolation procedures as approved by the Official State Agency;

(3) Eliminate other domesticated fowl from breeder farm;

(4) Dispose of all dead birds by burning, deep burial, or by putting them into special disposal pits.

(b) Recommended procedures:

(1) Avoid the introduction of Mycoplasma gallisepticum infected poultry;

(2) Prevent indirect transmission from outside sources through contaminated equipment, footwear, clothing, vehicles, or other mechanical means;

(3) Provide adequate isolation of breeder flocks to avoid airborne transmission from infected flocks;

(4) Minimize contact of breeder flocks with free-flying birds;

(5) Keep the rodent population and other pests under control;

(6) Tailor vaccination programs to needs of farm and area;

(7) Clean and disinfect equipment after each use;

(8) Provide clean footwear and provide an adequate security program;

(9) Clean and disinfect houses before introducing a new flock;

(10) Use well-drained range;

(11) Use clean, dry litter free of mold;

(12) Keep accurate records of death losses;

(13) Seek services of veterinary diagnostician if abnormal losses or signs of disease occur;

(14) Adopt and maintain a clean-egg program.

§ 147.27 Procedures recommended to prevent the spread of disease by artificial insemination of turkeys.

(a) The vehicle transporting the insemination crew shall be left as far as practical from the turkey pens.

(b) The personnel of the insemination crew should observe personal cleanliness, including the following sanitary procedures:

(1) Outer clothing shall be changed between visits to different premises so that clean clothing is worn upon entering each premises. The used apparel shall be kept separate until laundered. This also applies to gloves worn while handling turkeys;

(2) Boots or footwear shall be cleaned and disinfected between visits to different premises;

(3) Disposable caps shall be provided and discarded after use on each premises.

(c) The use of individual straw or similar technique is highly recommended. Insemination equipment which is to be reused shall be cleaned and disinfected before reusing. Equipment used for the convenience of the workers shall not be moved from premises to premises.

(d) No obviously diseased flock shall be inseminated. If evidence of active disease is noted after insemination is begun, operations should be stopped and the hatchery notified.

(e) Care should be taken during the collection of semen to prevent fecal contamination. If fecal material is present, it should be wiped off with a piece of cotton before the semen is collected. Likewise, care should be taken not to introduce fecal material into the oviduct of the hen.

Subpart D—Random Sample Performance Testing Procedures

§ 147.31 Random sample tests; general.

(a) The tests shall obtain specified performance data on representative samples of the stocks of two or more breeders, maintained under equal treatment with respect to housing, feeding, and management, at each test location.

(b) The tests shall be conducted by an impartial public agency.

(c) Samples shall be taken, preferably under the supervision of the Official State Agency, in accordance with the following procedures:

(1) The number and location of all flocks within the State supplying eggs of the grade to be tested shall be determined from Official State Agency records. By a process of drawing at random names or assigned numbers, determination shall be made from which of these flocks the sample is to be taken. The flock or flocks from which the sample is taken must include at least 1,000 birds.

(2) The eggs shall be taken from the nests, the farm egg room, or cases of hatching eggs or setting trays in the hatchery, in proportion to the number of birds in each flock represented.

(3) The sample shall not include eggs which, in the opinion of the sample taker, are unsuitable for hatching.

(4) The sample shall be placed in an appropriate container, and the container sealed with a distinctive seal or sealing tape by the sample taker.

(5) The sample taker shall furnish the Official State Agency and the test supervisor with a detailed report of the procedures followed in obtaining each sample.

(d) Entries shall be maintained in two or more replicates, and the performance of the replicates recorded separately.

(e) Pen assignments shall be made at random to reduce to a minimum any bias in results due to pen location.

§ 147.32 Random sample egg production tests.

(a) A minimum of 50 pullet chicks, hatched from the egg sample, shall be started for each entry.

(b) Records shall be kept on the performance of each entry until the birds reach 500 days of age.

(c) At the end of the test, and no later than November 1, the Supervisor shall submit to the ASR Division, for analysis and publication, a summary for each entry covering the following items:

(1) Name and address of entrant and the source of the sample;

(2) Breed or cross of breeds entered (indicating if entry is a pure strain, line cross, strain cross, breed cross, incross, incrossbred, or synthetic);

(3) Strain or trade name;

(4) Percent mortality to 150 days of age or subsequent age at housing;

(5) Percent laying house mortality computed from 150 days of age, or subsequent age at housing, to 500 days of age;

(6) Days of age to 50-percent production, calculated from the first day of the first 2 consecutive days of 50-percent production for living birds in the entry at that time;

(7) Number of eggs per pullet housed to 500 days of age;

(8) Percent hen-day production from the time the birds reached 50-percent production to 500 days of age;

(9) Income over feed and chick cost per pullet housed, with chick cost in 1,000 lots at hatch date adjusted for mortality (accidental deaths, sexing errors, and missing chicks not included);

(10) Pounds of feed per pound of eggs produced (weight of eggs produced shall be computed from production and egg weight records (bulk weighing) for each 2-week period throughout the test);

(11) Average annual egg weight, computed from bulk weighings at least every 2 weeks or 2 days a month at equal intervals;

(12) Percent large and extra large eggs, computed from all eggs laid 1 day each week per entry;

(13) Body weight at 150 days of age or subsequent age at housing, and at the end of test;

(14) Albumen quality—Haugh units measured on 1 day's eggs per quarter or every 3 months, at equal intervals, broken-out basis;

(15) Percentage of eggs with large blood spots, one-eighth inch or more, computed from at least 3 days' eggs per quarter, broken-out basis;

(16) Percentage of eggs with small blood spots, less than one-eighth inch, computed from at least 3 days' eggs per quarter, broken-out basis;

(17) Percentage of eggs with large colored meat spots, one-eighth inch or more, computed from at least 3 days' eggs per quarter, broken-out basis;

(18) Percentage of eggs with small colored meat spots, less than one-eighth inch, computed from at least 3 days' eggs per quarter, broken-out basis;

(19) Specific gravity score as determined from 1 day's eggs per quarter.

§ 147.33 Random sample meat production test.

(a) For the growing phase:

(1) An entry shall consist of at least 200 chicks hatched from a sample of eggs obtained as prescribed in § 147.31 or from an entry of the stock in the laying phase;

(2) Records shall be kept on the performance of each entry for a period determined by the test management;

(3) At the end of the test and no later than February 1, the Supervisor shall submit to the ASR Division, for analysis and publication, a summary for each entry covering the following items:

(i) Name and address of the entrant and the source of the sample;

(ii) Breed and strain or trade name of stock entered (including, for entries involving a cross of stocks, the identification of the stocks represented by the males and females in the parent flock);

(iii) Viability of chicks started to completion of test;

(iv) Average live weight of all pullets at completion of test;

(v) Average live weight of all cockerels at completion of test;

(vi) Percent eviscerated yield, by sexes, based on the live and eviscerated weights of all birds, or at least 50 birds of each sex selected at random, at the completion of the test;

(vii) Percent weight distribution in each U.S. Grade, by sexes, based on U.S. Classes, Standards and Grades for Poultry, as contained in 7 CFR Part 70, Subpart C (all factors considered except handling and dressing defects);

(viii) Feed conversion expressed as the pounds of feed required to produce a pound of live weight to the completion of test; and

(b) For the laying phase:

(1) An entry shall consist of a mating, including at least 50 pullets, representative of the stock entered. The birds in the entry shall be produced from a sample of eggs obtained as prescribed in § 147.31;

(2) Records shall be kept on the performance of each entry for a growing period of at least 150 days and an egg production period of 240 days;

(3) At the end of the test and no later than January 1, the Supervisor shall submit to the ASR Division, for analysis and publication, a summary for each entry covering the following items:

(i) Name and address of the entrant and the source of the sample;

(ii) Breed and strain or trade name of the stock entered and, for entries comprised of males of one and females of a different stock, the identification of each stock;

(iii) Percent mortality to 150 days of age or to subsequent age at housing;

(iv) Percent mortality from 150 days of age, or subsequent age at housing, to end of the 240-day period;

(v) Number of eggs per pullet housed to end of the 240-day period;

(vi) Percent hen-day production from the time the birds reached 50 percent production to end of the 240-day period. (The time of reaching 50 percent production shall be the first day of the first 2 consecutive days of 50 percent production for living birds in the entry at that time);

(vii) Average egg weight as computed from bulk weighings of all eggs laid at least 1 day a month;

(viii) Percent hatchability of all eggs set;

(ix) Body weight of females at end of test;

(x) Pounds of feed consumed during the 240-day period per dozen of eggs produced.

§ 147.34 Random sample tests; combined summary.

(a) The combined summary published by the ASR Division shall include the results of all acceptable tests, combined

by stocks, with adjustments for test differences, and the use of other accepted statistical procedures.

(b) The provisions specified in § 147.31 and either § 147.32 or § 147.33 shall be used by the ASR Division as a guide for determining acceptability of test results for inclusion in the combined summary.

Subpart E—Procedure for Changing National Poultry Improvement Plan

§ 147.41 Definitions.

Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:

(a) *Plan or NPIP.* The National Poultry Improvement Plan.

(b) *Plan Conference.* A meeting convened for the purpose of recommending changes in the provisions of the Plan.

(c) *Department.* The U.S. Department of Agriculture.

(d) *ASR Division.* The Animal Science Research Division of the Agricultural Research Service of the Department.

(e) *State.* Any State, the District of Columbia, or Puerto Rico.

(f) *Egg type chickens.* Chickens bred for the primary purpose of producing eggs for human consumption.

(g) *Meat type chickens.* Chickens bred for the primary purpose of producing meat.

(h) *Waterfowl.* Domesticated fowl that normally swim, such as ducks and geese.

(i) *Exhibition poultry.* Domesticated fowl which are bred for the combined purposes of meat or egg production and competitive showing.

(j) *Game birds.* Domesticated fowl, such as pheasants, partridge, quail, grouse, and guineas, but not doves and pigeons.

§ 147.42 General.

Changes in this subchapter shall be made in accordance with the procedure described in this subpart: *Provided,* That the Department reserves the right to make changes in this subchapter without observance of such procedure when such action is deemed necessary in the public interest.

§ 147.43 General Conference Committee.

(a) The General Conference Committee shall consist of the Poultry Coordinator in Charge, National Poultry Improvement Plan, ASR Division, and one member to be elected, as provided in paragraph (b) of this section, from each of the following regions:

(1) North Atlantic: Maine, New Hampshire, Vermont, Massachusetts, Rhode Island, Connecticut, New York, New Jersey, and Pennsylvania.

(2) East North Central: Ohio, Indiana, Illinois, Michigan, and Wisconsin.

(3) West North Central: Minnesota, Iowa, Missouri, North Dakota, South Dakota, Nebraska, and Kansas.

(4) South Atlantic: Delaware, District of Columbia, Maryland, Virginia, West Virginia, North Carolina, South Carolina, Georgia, Florida, and Puerto Rico.

(5) South Central: Kentucky, Tennessee, Alabama, Mississippi, Arkansas, Louisiana, Oklahoma, and Texas.

(6) Western: Montana, Idaho, Wyoming, Colorado, New Mexico, Arizona, Utah, Nevada, Washington, Oregon, California, Alaska, and Hawaii.

(b) The committee members will be elected by the official delegates of the respective regions. One alternate member shall also be elected from each region. There shall be at least two nominees for each position, and the voting shall be by secret ballot.

(c) Three members shall be elected at each NPIP Conference. Each member shall serve for a period of 4 years, subject to the continuation of the committee by the Secretary of Agriculture, and may not succeed himself.

(d) The duties of the General Conference Committee are as follows:

(1) Determine whether new proposals (i.e., proposals that have not been submitted as provided in § 147.44) may be considered. New proposals will be considered only with the unanimous consent of the committee.

(2) During the interim between conferences, the committee shall represent the cooperating States in:

(i) Reviewing and giving recommendations regarding the Department's report of changes and editing of this subchapter to include the changes.

(ii) Serving in an advisory capacity with respect to administrative procedures and interpretations of the provisions of this subchapter.

(iii) Recommending to the Secretary of Agriculture such administrative changes in the requirements of the Plan as may be necessitated by unforeseen conditions when postponement until the next conference would seriously impair the operation of the program. Such changes shall remain in effect only until confirmed or rejected by the next NPIP Conference, or until sooner rescinded by the committee;

(iv) Assisting the ASR Division in formulating plans for the next conference.

§ 147.44 Submitting, compiling, and distributing proposed changes.

(a) Changes in this subchapter may be proposed by any participant, Official State Agency, the Department, or other interested person or industry organization.

(b) Except as provided in § 147.43(d)(1), proposed changes shall be submitted in writing so as to reach the ASR Division not later than 150 days prior to the opening date of the Plan Conference, and participants in the Plan shall submit their proposed changes through their Official State Agency.

(c) The name of the proponent shall be indicated on each proposed change when submitted. Each proposal should be accompanied by a brief supporting statement.

(d) The ASR Division will notify all persons on the NPIP mailing lists concerning the dates and general procedure of the conference. Hatchery and dealer participants will be reminded of their privilege to submit proposed changes and to request copies of all the published proposed changes.

(e) The proposed changes, together with the names of the proponents and supporting statements, will be compiled by the ASR Division and issued in processed form. When two or more similar changes are submitted, the ASR Division will endeavor to unify them into one proposal acceptable to each proponent. Copies will be distributed to officials of the Official State Agencies cooperating in the NPIP. Additional copies will be made available for meeting individual requests.

§ 147.45 Official delegates.

Each cooperating State shall be entitled to one official delegate for each of the programs described in Subparts B, C, D, and E of Part 145 of this chapter in which it has one or more participants at the time of the conference. The official delegates shall be elected by a representative group of participating industry members and be certified by the Official State Agency. It is recommended but not required that the official delegates be Plan participants. Each official delegate shall endeavor to obtain, prior to the conference, the recommendations of industry members of his State with respect to each proposed change.

§ 147.46 Committee consideration of proposed changes.

(a) The following five committees shall be established to give preliminary consideration to the proposed changes falling in their respective fields:

(1) Egg Type Chickens.

(2) Meat Type Chickens.

(3) Turkeys.

(4) Waterfowl, Exhibition Poultry, and Game Birds.

(5) General and Auxiliary Provisions.

(b) Each official delegate shall be appointed a voting member in one of the committees specified in paragraph (a) of this section.

(c) Since several of the proposals may be interrelated, the committees shall consider them as they may relate to others, and feel free to discuss related proposals with other committees.

(d) The committees shall make recommendations to the conference as a whole concerning each proposal. The committee report shall show any proposed change in wording and the record of the vote on each proposal, and suggest an effective date for each proposal recommended for adoption. The individual committee reports shall be submitted to the chairman of the conference, who will combine them into one report showing, in numerical sequence, the committee recommendations on each proposal.

(e) The committee meetings shall be open to any interested person. Advocates for or against any proposal should feel free to appear before the appropriate committee and present their views.

§ 147.47 Conference consideration of proposed changes.

(a) The chairman of the conference shall be a representative of the Department.

(b) At the time designated for voting on proposed changes by the official delegates, the chairman of the General Conference Committee and the five committee chairmen shall sit at the speaker's table and assist the chairman of the conference.

(c) Each committee chairman shall present the proposals which his committee approves or recommends for adoption as follows: "Mr. Chairman. The committee on General and Auxiliary Provisions recommends the adoption of Proposal No. ------, for the following reasons (stating the reasons): I move the adoption of Proposal No. ------." A second will then be called for. If the recommendation is seconded, discussion and a formal vote will follow.

(d) Each committee chairman shall present the proposals which his committee does not approve as follows: "Mr. Chairman. The committee on General and Auxiliary Provisions does not approve Proposal No. ------." The chairman will then ask if any official delegate wishes to move for the adoption of the proposal. If moved and seconded, the proposal is subject to discussion and vote. If there is no motion for approval, or if moved but not seconded, there can be no discussion or vote.

(e) Discussion on any motion must be withheld until the motion has been properly seconded, except that the delegate making the motion is privileged, if he desires, to give reasons for his motion at the time of making it. To gain the floor for a motion or for discussion on a motion, the official delegate in the case of a motion, or anyone in case of discussion on a motion, shall rise, address the chair, give his name and State, and be recognized by the chair before proceeding further. While it is proper to accept motions only from official delegates and to limit voting only to such delegates, it is, however, equally proper to accept discussion from anyone interested. To conserve time, discussion should be pointed and limited to the pertinent features of the motion.

(f) Proposals that have not been submitted in accordance with § 147.44 will be considered by the conference only with the unanimous consent of the General Conference Committee. Any such proposals must be referred to the appropriate committee for consideration before being presented for action by the conference.

(g) Voting will be by States, and each official delegate, as determined by § 147.45, will be allowed one vote on each proposal pertaining to the program prescribed by the subpart which he represents.

(h) A roll call of States for a recorded vote will be used when requested by a delegate or at the discretion of the chairman.

(i) All motions on proposed changes shall be for adoption.

(j) Proposed changes shall be adopted by a majority vote of the official delegates present and voting.

(k) The conference shall be open to any interested person.

§ 147.48 Approval of conference recommendations by the Department.

Proposals adopted by the official delegates will be recommended to the Department for incorporation into the provisions of the NPIP. The Department reserves the right to approve or disapprove the recommendations of the conference as an integral part of its sponsorship of the National Poultry Improvement Plan.

Any person who wishes to submit written data, views, or arguments concerning the proposed revision of the National Poultry and Turkey Improvement Plans and Auxiliary Provisions may do so by filing them with the Director, Animal Science Research Division, Agricultural Research Center, Beltsville, MD 20705, within 30 days after publication hereof in the FEDERAL REGISTER.

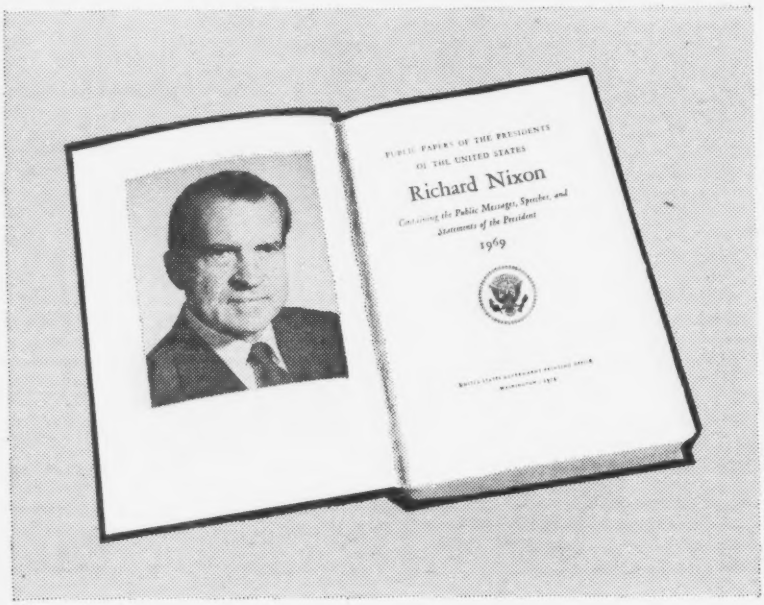
All written submissions made pursuant to this notice will be made available for public inspection at times and places and in a manner convenient to the public business (7 CFR 1.27(b)).

Done at Washington, D.C., this 11th day of May 1971.

GEORGE W. IRVING, Jr.,
Administrator,
Agricultural Research Service.

[FR Doc.71-6793 Filed 5-18-71;8:45 am]

PUBLIC PAPERS OF THE PRESIDENTS OF THE UNITED STATES



Richard Nixon — 1969

1183 Pages – Price: $14.50

CONTENTS

- Messages to the Congress
- Public speeches and letters
- The President's news conferences
- Radio and television reports to the American people
- Remarks to informal groups

PUBLISHED BY

Office of the Federal Register
National Archives and Records Service
General Services Administration

ORDER FROM

Superintendent of Documents
U.S. Government Printing Office
Washington, D.C. 20402

PRIOR VOLUMES

Volumes covering the administrations of Presidents Truman, Eisenhower, Kennedy, and Johnson are available at varying prices from the Superintendent of Documents, U.S. Government Printing Office, Washington, D.C. 20402.